# Detection of linezolid-resistant coagulase-negative staphylococci carrying cfr gene and PVL-positive S. simulans in environmental samples Laura Ruiz-Ripa<sup>1</sup>, Olouwafemi Mistourath Mama<sup>1</sup>, Susana Sanz<sup>1</sup>, Carmen Olarte<sup>1</sup>, Jose Manuel Azcona<sup>2</sup>, Carmen Torres<sup>1</sup>

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### Background

Linezolid is an important therapeutic option for the treatment of infections caused by multidrug-resistant (MDR) pathogens, including methicillin-resistant coagulase-negative staphylococci (MRCoNS)<sup>1</sup>. The cfr gene mediates transferable resistance to linezolid and it also confers resistance to phenicols, lincosamides, oxazolidinones, pleuromutilins and streptogramin A antimicrobials. Moreover, the Panton-Valentine leucocidin (PVL) is the most important toxin produced by Staphylococcus aureus, but there are few reports describing PVLproducing coagulase-negative staphylococci (CoNS)<sup>2</sup>.

Objective: To analyze the diversity of CoNS in environmental samples of swines farms, to characterize the antimicrobial resistance phenotype/genotype and to determine the presence of the PVL in recovered isolates.

### Methodology

Sampling and staphylococci recovery: In 2015, air and slurry pound samples of two Spanish pig farms, of intensive (Farm A) and semi-extensive (Farm B) production were collected. Samples were inoculated in mannitol salt agar (MSA) plates for staphylococci recovery and the species identification was performed by MALDI-TOF<sup>3</sup>.

**Antimicrobial resistance phenotype and genotype**: The susceptibility for 13 antimicrobials was determined by disk-diffusion method and the Minimum Inhibitory Concentration (MIC) to linezolid was performed by E-test (EUCAST, 2017). The presence of 32 antimicrobial resistance genes was determined by PCR and sequencing. In addition, mutations in the subunit GyrA were determined by PCR and sequencing in ciprofloxacin-resistant isolates<sup>4</sup>.

**Virulence factors**: The presence of the virulence factor *lukS/F-PV* enconding the PVL was likewise investigated by  $PCR^{5}$ .

### References

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## Aknowledgements

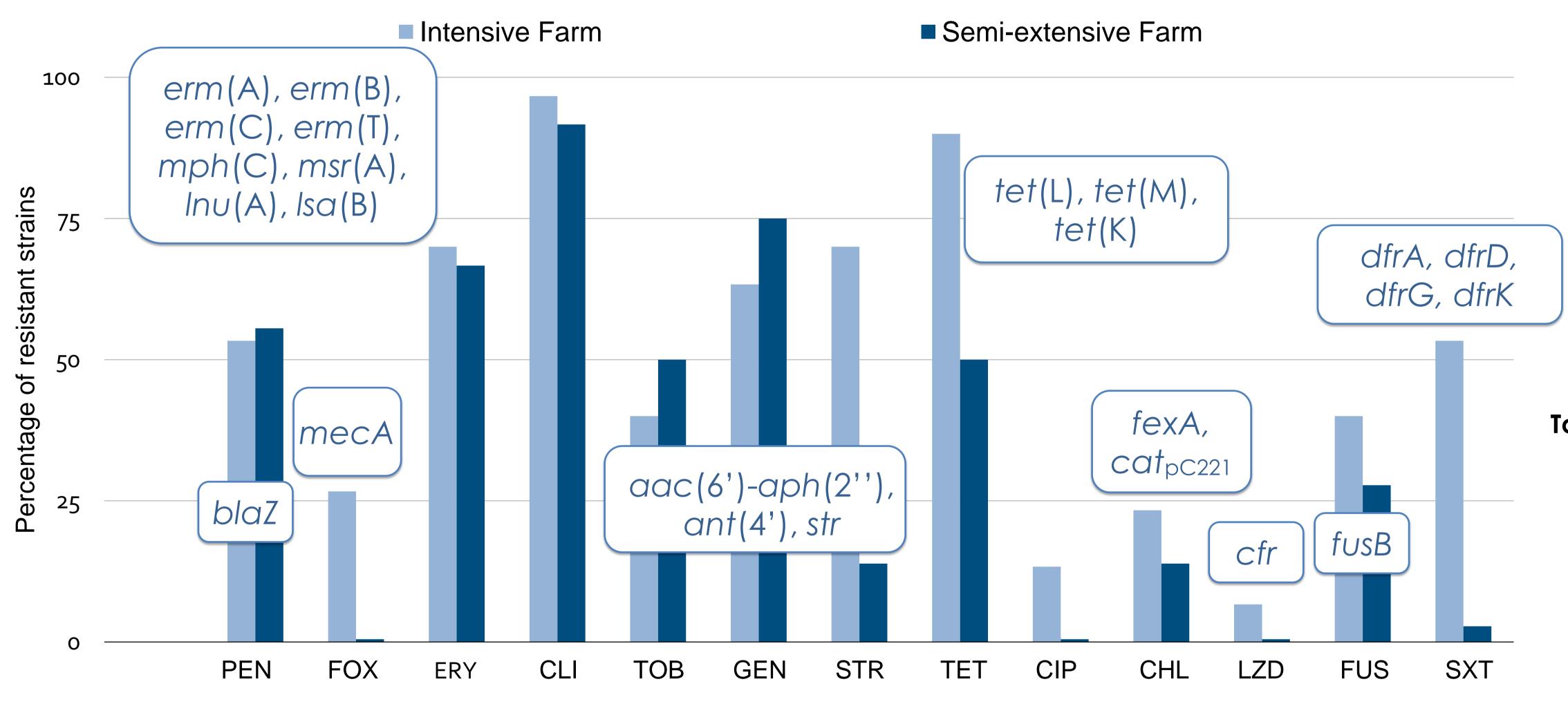
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67 CoNS isolates (thirty from Farm A and thirty-seven from Farm B) were recovered, belonging to 14 species with predominance of S. arlettae (n=18), S. cohnii (n=11), S. sciuri (n=9) and S. saprophyticus (n=5) (Figure 1). S. arlettae and S. cohnii were the species more frequently detected in the intensive and the semi-extensive farm, respectively.

Seven isolates (S. saprophyticus, 5; S. haemolyticus, 1; S. sciuri, 1) showed methicillin-resistance and harbored the mecA gene, being negative for mecB and mecC (Figure 2). Moreover, four isolates belonging to the S. sciuri group also carried the mecA gene but they did not show a methicillin-resistance phenotype. Only one isolate (S. chromogenes) was susceptible to all the antimicrobials tested and 70% of the strains were MDR. The percentage of resistant strains and the resistance genes detected are shown in Figure 2. Four ciprofloxacin-resistant isolates were detected, but only one *S. lentus* showed mutations in gyrA (S84L).

**Figure 2**. Percentage of resistant isolates depending on the swine farm and antimicrobial resistance genes detected.



PEN, penicillin; FOX, cefoxitin; ERY, erythromycin; CLI, clindamycin; TOB, tobramycin; GEN, gentamicin; STR, streptomycin; TET, tetracycline; CIP, ciprofloxacin; CHL, chloramphenicol; LZD, linezolid; FUS, fusidic acid; SXT, trimethoprim-sulfamethoxazole.

> 1. A high rate (70%) of multidrug-resistant CoNS was detected among environmental samples of swine farms. 2. The presence of cfr-positive linezolid-resistant CoNS in the air of pig farms, although this antimicrobial is not used in livestock, stressed the importance of coselection of resistances. 3. The detection of a PVL-positive S. simulans makes it necessary to monitor these virulent microorganisms in the environment.

- swine farms.

#### Results

### Conclusions

4. CoNS could act as a source of important antimicrobial resistance and virulence genes in environments with high selective pressure, like in the case of

Figure 1. N° of isolates of each species recovered from the swine fam. S. arlettae 🔳 S. cohnii 18 S. sciuri 24 S. saprophyticus Other species 11

Other species include S. haemolyticus, S. xylosus, S. epidermidis, S. equorum, S. nepalensis, S. warneri, S. lentus, S. chromogenes, S. simulans and S. succinus.

The two linezolid-resistant isolates S. equorum and S. arlettae (MIC=8 µg/ml) recovered from the farm with intensive production, carried the cfr gene and were negative for both optrA and poxtA genes (Table 1). The genes lukS/F-PV encoding the PVL were detected in one S. simulans isolate.

**Table 1.** Resistance genes detected in the llinezolid.resistant isolates.

Isolate	Resistance genes detected
S. equorum	cfr, mecA, lsa(B), erm(B), str, cat <sub>pC221</sub> , tet(M)
S. arlettae	cfr, lsa(B), mph(C), msr(A), lnua(A), str, tet(K),
	tet(M), fexA, dfrD



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