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# INTRA-ANNUAL TRENDS OF FUNGICIDES RESIDUES IN WATERS FROM VINEYARD AREAS IN LA RIOJA REGION OF NORTHERN SPAIN

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#### 1 Abstract

2 The temporal trends of fungicides in surface and ground water in ninety samples, 3 including both surface waters (12) and ground waters (78) from an extensive vineyard area 4 located in La Rioja (Spain) were examined between September 2010 and September 2011. 5 Fungicides are used in increasing amounts on vines in many countries, and they may reach the 6 water resources. However, few data have been published on fungicides in waters, with 7 herbicides being the more frequently monitored compounds. The presence, distribution and 8 year-long evolution of seventeen fungicides widely used in the region and a degradation 9 product were evaluated in waters during four sampling campaigns. All the fungicides 10 included in the study were detected at one or more of the points sampled during the four 11 campaigns. Metalaxyl, its metabolite CGA-92370, penconazole and tebuconazole were the 12 fungicides detected in the greatest number of samples, although myclobutanil, CGA-92370 13 and triadimenol were detected at the highest concentrations. The highest levels of individual fungicides were found in Rioja Alavesa, with concentrations of up to 25.52  $\mu$ g L<sup>-1</sup>, and more 14 than 40% of the samples recorded a total concentration of  $>0.5 \ \mu g \ L^{-1}$ . More than six 15 16 fungicides were positively identified in a third of the ground and surface waters in all the 17 sampling campaigns. There were no significant differences between the results obtained in the 18 four sampling campaigns, and corroborated a pattern of diffuse contamination from the use of 19 fungicides. The results confirm that natural waters in the study area are extremely vulnerable 20 to contamination by fungicides, and highlight the need to implement strategies to prevent and 21 control water contamination by these compounds.

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*Keywords*: multi-residue analysis; fungicides; surface water; groundwater; temporal
evaluation; vineyards.

#### 26 **1. Introduction**

27 In agriculture, the more intensive use of pesticides to prevent and combat fungi, weeds and 28 insects, the greater the potential discharge of these compounds into aquatic ecosystems. These 29 products or their degradation products, may remain in the environment, and this has become a 30 growing concern in recent years (Picó and Barceló, 2015; Ccanccapa et al., 2016; Teklu et al., 31 2016). Intensive agriculture combined with factors enhancing leaching and the hydrogeological characteristics of the unsaturated zone may lead to increased levels of 32 33 pesticides in groundwater (Vryzas et al., 2012). As a result of this, a persistent threat to the 34 environment and even to human health may occur, and it has required the drafting of strict directives by the European Commission (EC) to minimize the negative effects on the 35 environment. Council Directive 98/83/EC (EC, 1998) set the limits for pesticides in water 36 intended for human consumption at 0.1  $\mu$ g L<sup>-1</sup> for individual pesticides and 0.5  $\mu$ g L<sup>-1</sup> for the 37 38 sum of all pesticides. More recently, EC Directive (EC, 2000, 2008) established a framework 39 for Community action in the field of water policy, and approved a list of 33 priority 40 compounds in the field of water policy. One-third of the compounds included on this list are 41 pesticides, reflecting the EC's concern at the presence of these compounds in water (EC, 42 2013).

Fungicides are a group of pesticides used to combat fungi and (or) their spores. They can be chemical or biological, and act systemically or via contact with plant surfaces to prevent foliar diseases in a wide range of crops. The main market for fungicide is in Europe, where these compounds are critical components for the protection of grapevine and other crops (Oliver and Hewitt, 2014). According to data of FAOSTAT (2015) the use of fungicides and bactericides in the European Union was approximately 143500 t of active ingredients in 2013, which was higher than that in North America countries (85000 t of active ingredients).

A number of studies have reported fungicides in European waters. For example fungicides of the azole family have been found in samples from Swiss Midland lakes (Kahle et al., 2008); the metabolites of some fungicides (tolylfluanid, chlorothalonil, and trifloxystrobin) were found in waters from different areas in Germany (Reemtsma et al., 2013), and several fungicides were also detected by Gonçalves et al. (2007) in the waters of a vulnerable zone in Portugal, while carbendazim was the fungicide most frequently detected in several rivers and groundwaters influenced by agriculture in Serbia (Dujakovic et al., 2010).

57 Elsewhere azoxystrobin was the most frequently detected fungicide, followed by by metalaxyl, propiconazole, myclobutanil, and tebuconazole in southern and central USA 58 59 (Battaglin et al. 2011). Also in the US, Reilly et al. (2012) detected at least one fungicide in 75% and 58% of the surface and ground waters, respectively, from three geographical areas of 60 61 intense fungicide use. In Vietnam, isoprothiolane is the one most frequently detected in 62 samples from locations influenced by rice growing (Van Toan et al., 2013), in Costa Rica it 63 has been found in waters from an area impacted by nearby pineapple and banana plantations 64 (Echeverria-Saenz et al., 2012), and in Australia it was shown the presence of residues of 65 many different organic fungicides in the surface waters of a horticultural production 66 catchment (Wightwick et al. 2012).

67 In Spain, the occurrence of a relatively low number of fungicides waters in different 68 areas around the Mar Menor lagoon (SE Spain) (Moreno-González et al., 2013), in Andalusia 69 (southern Spain) (Belmonte Vega et al., 2005) and in the Ebro River basin (Ccanccapa et al., 2016) have been found. Fungicides were also detected in soils from the La Rioja (northern, 70 71 Spain), mainly metalaxyl and its metabolite, kresoxim-methyl and triadimenol (Pose-Juan et 72 al., 2015). Metalaxyl, cyprodinil and penconazole were detected in the sediments and soils in 73 different parts of Spain, such as a small river basin partially occupied by vineyards in Galicia (Bermúdez-Couso et al., 2007). Tebuconazole was detected in the basin of the River 74

Llobregat (Catalonia, Spain), although pesticide residues do not seem to pose a high risk to the local biota (Masiá et al., 2015). However, to date, there is clearly a lack of data regarding the presence and temporal evolution of fungicides in natural waters, especially in areas where they are widely used, such as agricultural zones where the main crop is vine.

79 Spain is the European country with the most land set aside for vineyards (14% of the 80 total area), and is the third in terms of wine production, behind Italy and France (International Organisation of Vine and Wine - OIV, 2015). La Rioja is the fifth region in Spain in grape 81 82 production. Its wines are protected by the oldest Qualified Designation of Origin (DOCa) in 83 Spain. The DOCa Rioja is the Spanish market leader for fine wines, with nearly 40% of the 84 total sales (Rioja DOCa - Qualified Designation of Origin, 2015). Hildebrandt et al. (2008) 85 and Navarro et al. (2010) reported the presence of different pesticides (including fungicides) 86 along the River Ebro, with some sampling points in La Rioja itself, although the monitoring 87 was too sporadic to provide a clear picture of water conditions in this area. Previous studies 88 we have carried out in this area have revealed the presence of a wide range of fungicides in surface and ground waters (Herrero-Hernández et al., 2012 and 2013). However, no study of 89 90 the temporal variation of fungicides in surface and ground waters has been conducted, 91 although one is required, considering the seasonal application of these compounds in 92 vineyards and to other crops in this area.

The aim of this work was to study the intra-annual trends of fungicides widely used in vine cultivation in ground and surface waters from the wine-growing region of La Rioja (Spain), and evaluate the pollution of natural waters by these compounds according to the levels permitted by EU legislation  $(0.1 \ \mu g \ L^{-1})$  (EC, 1998). Ninety sampling points, including wells, springs, uptakes and rivers, were sampled in the vineyard region of DOCa Rioja, and 17 fungicides were monitored in samples collected in four campaigns over the course of a year in order to explore the following: i) The temporal trends of the individual concentrations of fungicides in waters, ii) the temporal trends of the total concentrations of fungicides in
waters, and iii) the variability of the fungicide concentrations in all the samples from different
subareas and sampling campaigns.

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#### 104 **2. Materials and methods**

105 *2.1. Chemicals* 

The standards of 17 fungicides and a degradation product were supplied by Fluka, Dr.
Ehrenstorfer (Augsburg, Germany), Syngenta (Basel, Switzerland), and Riedel-de Haën
(Seelze-Hannover, Germany) (minimum purity higher than 98%). The common names and
physicochemical properties of the fungicides are listed in Table S1 in the Supplementary
Material.

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#### 112 2.2. Characterization of the study area

113 The DOCa Rioja wine region is located in northern Spain, straddling the River Ebro 114 (Fig. 1). The valley has a maximum width of around 40 km and an area of ~ 64,000 ha. It is 115 covered by vineyards with the DOCa designation that occupy successive terraces to an 116 altitude of around 700 m above sea level, and produce 280-300 million litres of wine per year. 117 This area is divided into three subareas: Rioja Alta (ALT), with an Atlantic climate and 118 chalky-clay, ferrous-clay or alluvial soils; Rioja Alavesa (ALV), with an Atlantic climate, and 119 chalky-clay soils on terraces and small plots, and Rioja Baja (BJ), with a drier and warmer 120 climate and alluvial and ferrous-clay soils (Herrero-Hernández et al., 2013). The region 121 usually records mild temperatures between 7°C and 20°C and an annual rainfall between 300 122 and 400 mm (Rioja DOCa - Qualified Designation of Origin, 2015) with a rainfall pattern 123 mainly winter dominated. The mobility of pesticides is favoured in the area because the soils

generally have a low organic matter (OM) content (<1.5 %) and a sandy clay loam or sandy</li>
loam texture (Pose-Juan et al., 2015).

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#### 127 2.3. Sample collection and analytical methodology

128 A total of ninety water samples were collected from three different agricultural areas 129 in the three different subareas of Rioja Alavesa (15 points), Rioja Alta (34 points), and Rioja 130 Baja (41 points). Twelve samples were surface waters (two on the River Ebro at the end of La 131 Rioja region, six on the main tributaries, one on the Lodosa canal, and three on small rivers), 132 and seventy-eight samples were groundwaters from private wells (44) with depths <10 m (39) 133 or >10 m (5), and uptakes or springs (Herrero-Hernández et al., 2013). Table S2 in the 134 Supplementary Material provides the characteristics of the sampling points in terms of water 135 type (surface or ground water) and water depth (well, spring, uptake or river), crop cultivated 136 and watering use. The wells were located inside the cultivated fields or next to them, and they 137 were generally used for irrigation purposes. Samples were collected manually or after 10 min of water pumping, using the pipes and the electrical pumps used by the farmer in each case. 138 139 They were collected over a year in four sampling campaigns: September 2010 (Sept-10), 140 March 2011 (Mar-11), June 2011 (Jun-11) and September 2011 (Sept-11).

141 All the water samples were collected in 2 L brown glass bottles and transported to the 142 laboratory in iceboxes. They were filtered through nitrocellulose screens with Minisart NY 25 143 filter 0.45 µm (Sartorius Stedim Biotech, Germany), and fungicides were removed by solid-144 phase extraction after a SPE preconcentration step with polymeric cartridges (Oasis HLB, 60 145 mg, Waters). Samples analysis was carried out by gas chromatography-mass spectrometry 146 (GC-MS) or by liquid-chromatography-mass spectrometry (LC-MS), as indicated in Herrero-147 Hernandez et al. (2012 and 2013). The analytical conditions and quality control parameters 148 are shown in Tables S3 and S4, respectively, in the Supplementary Material.

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2.4. Statistical analysis and data processing

151 Pearson correlations were calculated to relate the concentrations of fungicides in 152 waters and the properties of fungicides. Principal component analysis (PCA) was used as a 153 tool to explore the data variability of the fungicide concentrations in all the samples from 154 different subareas and sampling times. This multivariate data analysis transforms the set of 155 possible correlated measured variables into a reduced set of a few uncorrelated variables, and 156 it is used to facilitate the assessment of the potential contamination sources from data 157 obtained in large-scale monitoring studies. A total of 6480 results were evaluated from the 158 analysis of 360 samples. The matrix of scores and the matrix of loadings were obtained to 159 give information, respectively, on the distribution of patterns or sources of contamination 160 among samples (map of samples) and on the contribution of the original variables to each one 161 of these contamination patterns or sources (map of variables). IBM SPSS Statistics 22 162 software was used for data processing.

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165 3. Results and discussion

166 3.1. Intra-annual evolution of the individual concentrations of fungicides in waters in DOCa 167 Rioja

168 Residual concentrations of fungicides were detected in most of the samples analysed in the three subareas of DOCa Rioja (Tables 1-4), although there were several points where 169 170 fungicides were detected in any of the sampling periods. All the fungicides were detected in 171 one or more of the four sampling campaigns, and most of them were detected in one or more 172 of the samples; only iprovalicarb and cyproconazole were not detected in any sample in 173 September 2010 and March 2011, respectively. Other compounds were detected in five or

174 fewer samples, such as nuarimol, trifloxystrobin and dimethomorph in March 2011, nuarimol and trifloxystrobin in June 2011, and iprovalicarb, azoxystrobin and dimethomorph in 175 176 September 2011 (See Tables 1-4). These results confirm how vulnerable the natural waters in 177 the study area are to contamination by fungicides due to their widespread use in vineyards -178 the main crop. There was, nonetheless, a high percentage of sampling points that met the EU quality standard for individual compounds, with concentration levels below 0.1  $\mu$ g L<sup>-1</sup> 179 180 according to the percentages of total samples in which the fungicides were not detected, or 181 were detected below or above this legally established limit (See Fig. 2)

182 The most abundant and ubiquitous compounds detected in the first sampling campaign (Sept-10) were metalaxyl (52 samples, 12 of them with concentrations > 0.1  $\mu$ g L<sup>-1</sup> (52/12) 183 and a maximum concentration (Cmax) of 1.299  $\mu$ g L<sup>-1</sup>), its metabolite CGA-92370 (51/8, and 184 Cmax of 16.47  $\mu$ g L<sup>-1</sup>), penconazole (43/13 and Cmax of 0.342  $\mu$ g L<sup>-1</sup>), tebuconazole (42/14 185 and Cmax of 1.968  $\mu$ g L<sup>-1</sup>) and kresoxim-methyl (40/16 and Cmax of 0.287  $\mu$ g L<sup>-1</sup>). Other 186 187 compounds detected in this sampling in high concentrations were myclobutanyl, azoxystrobin and triadimenol, at Cmax of 12.61, 4.778 and 4.332  $\mu$ g L<sup>-1</sup> respectively. In all cases, these 188 189 Cmax were detected in samples from Rioja Alavesa.

Some of these compounds, such as tebuconazole (67/3 and Cmax of 3.236  $\mu$ g L<sup>-1</sup>), 190 metalaxyl (45/12 and Cmax of 8.015  $\mu$ g L<sup>-1</sup>), kresoxim-methyl (45/7 and Cmax of 0.574  $\mu$ g 191  $L^{-1}$ ), and penconazole (41/5 and Cmax of 3.683 µg  $L^{-1}$ ), were also the most ubiquitous 192 193 compounds in the sampling campaign carried out in Mar-11, together with pyrimethanil (58/6 and Cmax of 0.234  $\mu$ g L<sup>-1</sup>) and triadimenol (40/12 and Cmax of 3.103  $\mu$ g L<sup>-1</sup>). The metabolite 194 of metalaxyl and myclobutanyl were the other compounds detected in high concentrations 195 (Cmax of 7.226 and 7.208  $\mu$ g L<sup>-1</sup>, respectively). Cmax data were recorded for tebuconazole, 196 197 myclobutanil, triadimenol, metalaxyl and its metabolite in Rioja Alavesa, for penconazole, 198 triadimenol and metalaxyl in Rioja Alta, and for metalaxyl and triadimenol in Rioja Baja.

199 The same fungicides were also detected in the sampling campaigns carried out in Jun-11 (metalaxyl (59/19 and Cmax of 18.85  $\mu$ g L<sup>-1</sup>), its metabolite CGA-92370 (55/9 and Cmax 200 of 1.643  $\mu$ g L<sup>-1</sup>), and triadimenol (34/14 and Cmax of 8.470  $\mu$ g L<sup>-1</sup>), together with 201 azoxystrobin (50/3 and Cmax of 0.463  $\mu$ g L<sup>-1</sup>) and myclobutanil (33/18 and Cmax of 3.091 202  $\mu$ g L<sup>-1</sup>); and in the last sampling campaign carried out in Sept-11, penconazole (47/12 and 203 Cmax of 0.476  $\mu$ g L<sup>-1</sup>), metalaxyl (39/8 and Cmax of 1.848  $\mu$ g L<sup>-1</sup>), cyproconazole (39/4 and 204 Cmax of 0.154  $\mu$ g L<sup>-1</sup>), tebuconazole (37/7 and Cmax of 3.023  $\mu$ g L<sup>-1</sup>) and myclobutanil (33/6 205 and Cmax of 5.068  $\mu$ g L<sup>-1</sup>). High concentrations were also detected in Rioja Alavesa and in 206 207 Rioja Alta in the June-11 and Sep-11 sampling campaigns (Tables 1-4).

208 It is evident that metalaxyl, its metabolite CGA-92370, penconazole and tebuconazole 209 were the fungicides detected in a greater number of samples, although myclobutanil, CGA-210 92370 and triadimenol were detected in higher concentrations. Fig. 3 shows the evolution of 211 the sum of concentrations of these compounds in all the samples over the four campaigns in 212 the three subareas of DOCa Rioja. Peak concentrations were found in March (penconazole 213 and tebuconazole), June (metalaxyl and myclobutanil), and September (CGA-92370, myclobutanil, triadimenol and tebuconazole), and they could be related to the application of 214 215 these fungicides following the recommendations made by the authorities and experts for 216 tackling ad hoc diseases in the different areas (Gobierno de La Rioja, 2016). The highest 217 fungicide concentrations were obtained for samples located in Rioja Alavesa, except for 218 penconazole, which recorded a concentration peak in Rioja Alta.

A certain relationship was found between water solubility, GUS index or Kow, and the total concentrations of these fungicides, although it was significant only for water solubility (excluding metalaxyl) in the June sampling (r=0.88, p<0.10), for the GUS index in the September sampling (r=0.82, p<0.10), and for Kow (r=-0.94 and -0.99 (p<0.01)) in the March and June samplings, respectively. However, no significant correlations were found for theseparameters when all the fungicides were considered jointly.

225 Residues of the fungicides included in this study were also detected in surface waters 226 in the province of Almería (SE Spain) (pyrimethanil, myclobutanil, benalaxyl, nuarimol, 227 tebuconazole and azoxystrobin) (Ruiz Gil et al., 2008). Furthermore, the levels of fungicides 228 found are similar to those reported in other European agricultural regions in Germany 229 (Berenzen et al., 2005) and Greece, where triadimenol, cyproconazole, pyrimethanil and 230 benalaxyl were detected in several water samples (Thomatou et al., 2013), and more recently, 231 fungicides have been detected in 14% of the samples in the basin of Lake Vistonis, with 232 metalaxyl being the compound found in higher concentrations (Papadakis et al., 2015).

233 Other studies found residues of some of the fungicides included in this work in several 234 agricultural regions in countries as Argentina (tebuconazole with a mean concentration of 0.033 µg L<sup>-1</sup>) (De Geronimo et al., 2014), Costa Rica (myclobutanil, triadimenol, cyprodinil 235 and metalaxyl reaching values of 0.082  $\mu$ g L<sup>-1</sup> for metalaxyl ) (Echeverría-Sáenz et al., 2012), 236 Brazil (tebuconazole, azoxystrobin and trifloxystrobin reaching concentrations up to 0.037 µg 237 L<sup>-1</sup> in river water samples) (Montagner et al., 2014), USA in samples from streams, ponds, 238 239 and shallow groundwaters in areas of intense fungicide use (azoxystrobin, pyrimethanil, cyprodinil and dimethomorph with concentrations up to 0.180  $\mu$ g L<sup>-1</sup> (Reilly et al., 2012) and 240 in water samples from streams in 13 states (azoxystrobin, metalaxyl, myclobutanil, 241 tebuconazole and trifloxystrobin with concentrations up to 1.13  $\mu$ g L<sup>-1</sup>) (Battaglin et al., 242 2011), and Australis in the surface water samples (myclobutanil, pyrimethanil, cyproconazole, 243 trifloxystrobin, and fenarimol at concentrations  $>0.2 \ \mu g \ L^{-1}$ ) (Wightwick et al. 2012). 244

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246 3.2. Intra-annual evolution of the total concentration of fungicides in waters in DOCa Rioja

247 The temporal evolution of the total concentration of fungicides was studied to 248 establish the 'good chemical status' of surface and ground waters according to the European Directive (EC, 2008). This Directive establishes a limit of 0.5  $\mu$ g L<sup>-1</sup> for the aggregate 249 250 concentration of pesticides to be applied to drinking water. In groundwater (Fig. 4), the results 251 in Sept-10 showed up to 11 samples in Rioja Alta (35%) with  $\sum$  concentration of fungicides above 0.5 µg L<sup>-1</sup>, and a number of samples in Rioja Alavesa (2) and Rioja Baja (1) with 252  $\Sigma$  concentrations above 25 µg L<sup>-1</sup>. In Mar-11, up to nine samples were detected in Rioja Alta 253 (29%) with  $\Sigma$  concentration of fungicides above 0.5 µg L<sup>-1</sup>, as well as samples with 254 concentrations >15  $\mu$ g L<sup>-1</sup> in Rioja Alavesa (2), and >10  $\mu$ g L<sup>-1</sup> in Rioja Alta (1) and Rioja 255 256 Baja (1). In Jun-11, up to nine samples were detected in Rioja Alta (29%) containing fungicide concentrations above 0.5  $\mu$ g L<sup>-1</sup>, and samples with concentrations above 8  $\mu$ g L<sup>-1</sup> 257 258 were found in Rioja Alavesa (3) and Rioja Baja (1). In the last sampling campaign (Sept-11), 259 up to five samples were found in Rioja Baja (17%) containing fungicides adding up to more than 0.5  $\mu$ g L<sup>-1</sup>, and there were three samples in Rioja Alavesa with concentrations above 260 9  $\mu$ g L<sup>-1</sup>. 261

262 No significant relationship was found between the depth of the groundwater and the 263 total content of fungicides. The wells deeper than 10 m recorded, in general, the lowest 264 content of fungicides although it was not possible to find relationships with parameters of 265 soils or local hydrologic conditions. However it was observed that less polluted groundwaters 266 corresponded to wells located in areas where the main crops are fruit trees, cereals, etc. with 267 lower amounts of fungicides than in vineyard are applied. On the other hand it worth noting 268 that the high levels of total concentration over the four sampling campaigns in Rioja Alavesa 269 (2), Rioja Alta (1) and Rioja Baja (1) were found at the same sampling points. They 270 correspond to groundwater samples from shallow wells (1-3 m) in areas where the vineyard is the only crop and this may be due to improper handling of these products in the vicinity ofwells.

In surface waters, the results obtained in the four sampling periods indicated  $\sum$  concentrations of fungicides above 0.5 µg L<sup>-1</sup> in 0-2 samples (0-66%) in Rioja Alavesa, in 2-3 samples (33-100%) in Rioja Alta, and in 1-2 samples (16-66%) in Rioja Baja.

This total concentration of fungicides in waters is generally due to the presence of a 276 277 different number of compounds. In groundwaters, the results indicate that no fungicides were 278 detected in just seven of the 78 samples in Sept-10, eight in Mar-11, two in Jun-11 and 279 seventeen in Sept-11, but more than six fungicides were detected in a higher number of 280 samples (29 in Sept-10, 23 in Mar-11, 25 in Jun-11, and 23 in Sept-11) (Fig 5a). In surface 281 waters, the number of samples positively identifying more than six fungicides was four in 282 Sept-10, five in Mar-11 and Jun-11, and six in Sept-11 (Fig 5b). The percentage of samples 283 with a high number of fungicides (29-37% of groundwaters and 33-50% of surface waters) 284 again highlights the widespread use of different compounds on local crops (vines and fruits) 285 due to the different diseases affecting them, which are mainly Downy mildew (Plasmapora 286 viticola), Botrytis cinerea, and Powdery mildew (Uncinula necator).

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#### 288 3.3. Principal component analysis

Principal component analysis (PCA) was used as a multivariate analysis tool to explore the relationships among the concentrations of fungicides measured in different subareas and sampling campaigns, and evaluate their variability. The results indicate that all the variables are explained to a greater or lesser extent by the factors obtained from the analysis. In agreement with the explained total variance, the number of principal components (PCs) with eigenvalues >1 that better explain the variables were six and five, respectively, for ground and surface waters. These PCs explain 70.93% of the variance of fungicide data from groundwater samples, whereas a higher variance (73.79%) was explained by the data from
surface water samples. Similar variances were explained when data were arranged by subarea
or sampling campaign.

299 For groundwater, the first PC explained 25.38% of the total data variance, and for 300 surface waters this first PC explained 25.71% of the variance (Table 5). In general, all the 301 loadings were positive and high, indicating a pattern of diffuse contamination from the use of 302 fungicides in the areas studied. This first PC was mainly explained for groundwaters by the 303 presence of myclobutanil, tebuconazole and dimethomorph, and for surface waters by the 304 presence of cyprodinil and pyrimethanil, while other fungicides made a lower contribution to 305 PC1. Certain variables, such as triadimenol, metalaxyl or its metabolite CGA-92370, recorded 306 a greater loading in other PCs (Table 5). The results indicate that CGA-92370 and metalaxyl 307 did not dominate similar PCs, although CGA-92370 is produced by the degradation of 308 metalaxyl.

309 Score plots for PC1 vs. PC2, representing the distribution of sampling sites from 310 different subareas and sampling campaigns corresponding to groundwaters (Fig. 6a) and 311 surface waters (Fig. 6b) as well as the distribution of sampling sites from different campaigns 312 corresponding to ground and surface waters (Fig. 7) were only included. Score plots for data 313 distributed in other limited regions of space encompassed by different well-defined PC axes 314 are not shown because they are not relevant. The distribution of variables in different 315 groundwaters was fairly uniform around the origin of the axis, although some sites in Rioja 316 Alavesa and Rioja Alta are outstanding (Fig. 6a). The variables from Rioja Alavesa 317 groundwaters stand out in all the sampling campaigns, whereas the variables from Rioja Alta 318 groundwaters only do so in the Mar-11 sampling. This agrees with the high values found for 319 metalaxyl, CGA-92370, tebuconazole, triadimenol and myclobutanil in some sites in Rioja Alavesa, and penconazole in some sites in Rioja Alta. The outstanding points in the Mar-11 320

321 sampling might indicate residual contamination from previous campaigns at certain hot spots.
322 The variables were also uniformly distributed around the origin of the axis, with some sites
323 standing out in different surface waters. In this case, Rioja Alta sites recorded greater
324 variability in the four sampling campaigns, whereas they were only outstanding for Rioja
325 Alavesa in the Jun-11 sampling.

326 The variables are distributed around the origin of the axis in different surface waters, 327 whereas they are distributed in the lower quadrant with only some outstanding points through 328 the upper right quadrant in groundwaters in all the sampling campaigns (Fig. 7). Surface 329 water recorded lower levels of fungicides than groundwaters and a lower detection rate. These 330 results could be explained by the behavior (sorption and leaching) of fungicides in soils which 331 is depending on the characteristics of soils and fungicides. In a previous study a significant 332 correlation was found between residues of some fungicides in soils from Rioja region and soil 333 characteristics (Pose-Juan et al., 2015). This finding was also reported in other studies on the 334 distribution of pesticides in other areas (Hildebrandt et al., 2008), and it was explained by the 335 rapid degradation of pesticides in surface water, whereas a lower degradation of these 336 compounds could occur in groundwater, making this medium more vulnerable, especially 337 when groundwater is used for human consumption.

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#### **4. Conclusions**

The study carried out between September 2010 and September 2011 along the wine growing region of La Rioja showed the great diversity of fungicides in the study area and its occurrence in water samples with a dispersed pattern of fungicide concentrations. All the fungicides included in the study were detected in one or more of the four sampling campaigns, and most of them were detected in one or more of the samples in all the campaigns. Metalaxyl, its metabolite CGA-92370, penconazole and tebuconazole were the

most frequently detected fungicides, although myclobutanil, CGA-92370 and triadimenol 346 347 were detected in higher concentrations. These results are consistent with the widespread use 348 of these compounds which are those more applied following information provided by local 349 farmers. The highest concentrations of fungicides were obtained in samples located in Rioja 350 Alavesa regard other zones of the study area, except penconazole with the highest total 351 concentration located in Rioja Alta. Up to 40%, 35% and 25% of groundwaters with a total concentration of fungicides  $>0.5 \ \mu g \ L^{-1}$  were detected in the areas studied, and approximately 352 353 a quarter of the total waters recorded more than six fungicides in all the sampling campaigns. 354 Furthermore, the concentrations in groundwaters were greater than in surface waters in all 355 cases, and samples not too deep located close to vineyard showed the higher concentrations. 356 This could indicate an easier percolation of fungicides in these cases and highlight the need to 357 implement strategies to enhance the retention of these compounds by soils for effective water 358 protection given the imperative use of these compounds in agriculture. The use of biobeds 359 during the handling of pesticides or the washing of equipment and the application of organic 360 amendments to soils may be some of these strategies.

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483 **Figure captions** 

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485 Figure 1. Map of the Rioja DOCa wine-growing region in Spain including the sampling
486 points (taken from "Plan Nacional de ortofotografía aérea de España").

- 487 Figure 2. Distribution of samples with no detected fungicides or detected in concentrations
- 488 below or over the limit established by European Directives for human consumption (0.1 μg L<sup>-</sup>
- <sup>1</sup>) in different sampling campaigns: September 2010 (a), March 2011 (b), June 2011 (c) and
  September 2011 (d).
- 491 Figure 3. Changes of total concentrations of the most frequently detected fungicides:
  492 metalaxyl, its metabolite CGA-92370, myclobutanil, triadimenol, penconazole and
  493 tebuconazole detected in all sampling sites of the different subareas of La Rioja.
- 494 **Figure 4.** Distribution of ground (a) and surface water (b) samples with total concentration of 495 fungicides below or over the limit established by European Directives for human consumption 496  $(0.5 \ \mu g \ L^{-1})$  in different sampling campaigns in the three subareas of La Rioja.
- 497 Figure 5. Percentage of ground (a) and surface water (b) samples with different number of498 fungicides detected for the four sampling campaigns in the three subareas of La Rioja.
- 499 Figure 6. Scores plot for PC1 vs. PC2 showing data distribution of ground waters (a) and
- 500 surface waters (b) for all the sampling campaigns in the subareas of Rioja Alavesa (ALV),
- 501 Rioja Alta (ALT) and Rioja Baja (BJ).
- Figure 7. Scores plot for PC1 vs. PC2 showing data distribution depending on the type of
  water for all the sampling campaigns in the three subareas of La Rioja (GW, groundwater;
  SW, surface water).

Rioja Alavesa (n = 15)Rioja Alta (n = 34)Rioja Baja (n = 41)Concentration Concentration Concentration Pesticide Positive samples Positive samples Positive samples  $(\mu g L^{-1})$  $(\mu g L^{-1})$  $(\mu g L^{-1})$ C < 0.1 C > 0.1Average±SD  $C_{max}$ C < 0.1 C > 0.1 Average±SD  $C_{max}$ C < 0.1 C > 0.1Average±SD  $C_{max}$ Cymoxanil 0 0 0 0 0.257±0.193 0.449 4 2  $0.089 \pm 0.073$ 0.222 2 1 Pyrimethanil 3 3 0.125±0.107 0.331 8 8 0.167±0.167 0.590 10 4  $0.089 \pm 0.130$ 0.478 Cyprodinil 2 2 0.109±0.104 0.124±0.109 3 0.126±0.087 0.249 5 4 0.352 2 0.247 Iprovalicarb 0 0 0 0 \_ 0 0 \_ \_ --Metalaxyl 8  $0.278 \pm 0.480$  $0.105 \pm 0.160$ 0.590  $0.051 \pm 0.099$ 4 1.299 15 7 17 1 0.435 CGA 92370 7 5  $1.853 \pm 4.805$ 17 0.051±0.099 0.435 19 2  $0.062 \pm 0.148$ 0.637 16.47 1 0.095±0.116 7  $0.084 \pm 0.070$ 0.222 Penconazole 6 3 0.342 14 3  $0.064 \pm 0.066$ 0.209 10 Myclobutanil 2 2  $3.487 \pm 6.107$  $0.087 \pm 0.068$ 0.239  $0.162 \pm 0.232$ 0.750 12.61 7 3 5 4 Triadimenol 6 2 0.903±1.661 4.332 5 0 0.037±0.015 0.063 10 0  $0.034 \pm 0.012$ 0.045 2  $0.018 \pm 0.006$ 0.022  $0.034 \pm 0.012$ 0.045 0.021±0.012 0.042 Flutriafol 0 10 0 8 0 Tebuconazole  $0.086 \pm 0.041$ 4 4 0.321±0.667 1.968 11 4 0.164 13 6  $0.084 \pm 0.048$ 0.206 Kresoxim-methyl 4 3  $0.109 \pm 0.094$ 0.279 7 0.124±0.082 0.287 7  $0.078 \pm 0.071$ 0.206 6 13 Nuarimol 3  $0.100 \pm 0.088$ 1 0.162 11 4  $0.076 \pm 0.061$ 0.202 8 3  $0.064 \pm 0.048$ 0.167 2 0.214 Benalaxvl 1 1  $0.094 \pm 0.073$ 0.146 10  $0.089 \pm 0.091$ 0.325 4 6  $0.108 \pm 0.062$ Azoxyxtrobin  $0.878 \pm 1.911$ 0.209  $0.058 \pm 0.029$ 0.103 3 3 4.778 7 3  $0.085 \pm 0.058$ 7 1 Trifloxystrobin 0 7 4 0.037±0.028 0.077 0 0.053±0.015 0.075 10 0  $0.036 \pm 0.026$ 0.086 Dimethomorph 3 2  $0.067 \pm 0.071$ 0.170 10 0 0.036±0.019 0.075 6 0  $0.023 \pm 0.008$ 0.036 Cyproconazole 0.094 0.052±0.024 0.092  $0.049 \pm 0.025$ 0.102 0 0 1 12 14 1 -

Number of positive samples (or samples with some compound detected) with concentrations below and over 0.1  $\mu$ g L<sup>-1</sup> and average and maximum concentrations for the fungicides detected in the different areas of study in September 2010.

Number of positive samples (or samples with some compound detected) with concentrations below and over 0.1  $\mu$ g L<sup>-1</sup> and average and maximum concentrations for the fungicides detected in the different areas of study in March 2011.

		Rioja Ala	avesa (n = $15$ )			Rioja A	Alta $(n = 34)$			Rioja B	aja (n = 41)	
Pesticide	Positive samples		Concentra (µg L <sup>-1</sup>		Positive	samples	Concentra (µg L <sup>-1</sup>		Positive	samples	Concentra $(\mu g L^{-1})$	
	C < 0.1	C > 0.1	Average±SD	$C_{max}$	C < 0.1	C > 0.1	Average±SD	$C_{max}$	C < 0.1	C > 0.1	Average±SD	$C_{max}$
Cymoxanil	0	0	-	-	5	2	$0.189 \pm 0.315$	0.900	0	2	$0.166 \pm 0.075$	0.219
Pyrimethanil	3	3	$0.058 \pm 0.016$	0.089	22	2	$0.057 \pm 0.041$	0.234	27	1	$0.049 \pm 0.016$	0.105
Cyprodinil	2	0	$0.080 \pm 0.005$	0.084	5	4	0.159±0.219	0.737	5	0	$0.052 \pm 0.033$	0.084
Iprovalicarb	2	1	$0.077 \pm 0.030$	0.111	4	1	$0.107 \pm 0.118$	0.315	1	1	$0.147 \pm 0.080$	0.204
Metalaxyl	7	2	0.536±1.139	3.396	14	5	$0.204 \pm 0.610$	2.707	12	5	0.526±1.931	8.015
CGA 92370	3	4	$1.566 \pm 2.785$	7.226	9	4	$0.068 \pm 0.042$	0.164	8	4	$0.185 \pm 0.300$	1.089
Penconazole	9	0	$0.052 \pm 0.033$	0.089	15	4	$0.286 \pm 0.843$	3.683	12	1	$0.050 \pm 0.089$	0.338
Myclobutanil	3	5	$1.033 \pm 2.498$	7.208	4	9	0.189±0.137	0.474	6	7	$0.148 \pm 0.147$	0.404
Triadimenol	5	2	$0.573 \pm 0.965$	2.476	10	4	$0.274 \pm 0.755$	2.889	13	6	$0.277 \pm 0.698$	3.103
Flutriafol	7	1	$0.058 \pm 0.033$	0.100	7	1	$0.106 \pm 0.154$	0.484	4	0	$0.038 \pm 0.009$	0.046
Tebuconazole	13	1	$0.270 \pm 0.854$	3.236	26	0	$0.042 \pm 0.019$	0.093	25	2	$0.054 \pm 0.041$	0.205
Kresoxim-methyl	8	1	$0.078 \pm 0.054$	0.212	16	3	$0.088 \pm 0.122$	0.574	14	3	$0.075 \pm 0.047$	0.232
Nuarimol	2	0	0.071±0.025	0.089	1	0	0.034	0.034	2	0	$0.045 \pm 0.003$	0.048
Benalaxyl	2	0	$0.037 \pm 0.007$	0.043	3	0	$0.063 \pm 0.017$	0.081	3	0	$0.054 \pm 0.033$	0.091
Azoxystrobin	1	1	0.493±0.636	0.943	5	0	$0.042 \pm 0.023$	0.074	3	0	$0.042 \pm 0.025$	0.070
Trifloxystrobin	0	0	-	-	2	1	0.166±0.236	0.439	1	0	0.025	0.025
Dimethomorph	0	2	0.222±0.069	0.271	0	0	-	-	1	0	0.013	0.013
Cyproconazole	0	0	-	-	0	0	-	-	0	0	-	-

		Rioja Ala	avesa (n = 15)		Rioja A	Alta (n = 34)		Rioja Baja (n = 41)				
Pesticide	Positive samples		Concentra $(\mu g L^{-1})$		Positive	samples	Concentra (µg L <sup>-1</sup>		Positive	samples	Concentra (µg L <sup>-1</sup>	
	C < 0.1	C > 0.1	Average±SD	$C_{max}$	C < 0.1	C > 0.1	Average±SD	$C_{max}$	C < 0.1	C > 0.1	Average±SD	$C_{max}$
Cymoxanil	4	1	0.176±0.277	0.670	4	1	$0.083 \pm 0.048$	0.163	3	3	0.117±0.109	0.313
Pyrimethanil	3	0	$0.044 \pm 0.022$	0.068	7	1	$0.050 \pm 0.045$	0.143	3	2	$0.093 \pm 0.058$	0.169
Cyprodinil	6	0	$0.018 \pm 0.009$	0.029	13	0	$0.021 \pm 0.016$	0.051	7	0	$0.019 \pm 0.008$	0.028
Iprovalicarb	5	1	$0.094 \pm 0.021$	0.133	8	1	$0.076 \pm 0.027$	0.135	13	2	$0.074 \pm 0.021$	0.110
Metalaxyl	6	6	$2.194 \pm 5.416$	18.85	14	9	$0.117 \pm 0.132$	0.613	20	4	$0.169 \pm 0.351$	1.607
CGA 92370	8	3	$0.326 \pm 0.577$	1.643	20	2	$0.066 \pm 0.055$	0.230	18	4	$0.070 \pm 0.063$	0.245
Penconazole	3	4	$0.161 \pm 0.143$	0.429	6	5	$0.139 \pm 0.104$	0.336	8	5	$0.119 \pm 0.117$	0.363
Myclobutanil	0	6	$0.759 \pm 1.154$	3.091	1	9	$0.183 \pm 0.118$	0.374	14	3	$0.083 \pm 0.074$	0.307
Triadimenol	5	4	$0.762 \pm 1.690$	5.218	0	8	$1.218 \pm 2.930$	8.470	15	2	$0.139 \pm 0.322$	1.362
Flutriafol	3	0	$0.037 \pm 0.006$	0.042	6	0	$0.038 \pm 0.009$	0.054	8	0	0.035±0.013	0.054
Tebuconazole	7	1	$0.160 \pm 0.374$	1.086	5	0	$0.044 \pm 0.021$	0.079	7	1	$0.060 \pm 0.081$	0.254
Kresoxim-methyl	6	0	$0.053 \pm 0.022$	0.083	11	1	$0.066 \pm 0.024$	0.105	11	2	$0.089 \pm 0.108$	0.428
Nuarimol	1	1	0.102±0.066	0.149	2	0	$0.055 \pm 0.049$	0.089	0	1	0.114	0.114
Benalaxyl	4	1	$0.057 \pm 0.062$	0.167	8	2	$0.045 \pm 0.041$	0.135	8	1	$0.060 \pm 0.031$	0.125
Azoxystrobin	9	1	0.078±0.136	0.463	17	2	$0.046 \pm 0.051$	0.241	21	0	$0.030 \pm 0.012$	0.067
Trifloxystrobin	1	0	0.089	0.089	1	0	0.052	0.052	1	1	$0.103 \pm 0.085$	0.163
Dimethomorph	3	2	$0.087 \pm 0.108$	0.238	6	1	$0.028 \pm 0.036$	0.109	6	0	$0.019 \pm 0.022$	0.061
Cyproconazole	4	1	$0.200 \pm 0.343$	0.811	3	0	$0.046 \pm 0.017$	0.066	0	2	0.277±0.217	0.430

Number of positive samples (or samples with some compound detected) with concentrations below and over 0.1  $\mu$ g L<sup>-1</sup> and average and maximum concentrations for the fungicides detected in the different areas of study in June 2011.

Number of positive samples (or samples with some compound detected) with concentrations below and over 0.1  $\mu$ g L<sup>-1</sup> and average and maximum concentrations for the fungicides detected in the different areas of study in September 2011.

		Rioja Ala	avesa (n = 12)			Rioja A	Alta $(n = 30)$			Rioja E	Baja (n = 40)	
Pesticide	Positive samples		Concentra (µg L <sup>-1</sup>		Positive	samples	Concentra (µg L <sup>-1</sup>		Positive	samples	Concentra (µg L <sup>-1</sup>	
	C < 0.1	C > 0.1	Average±SD	$C_{max}$	C < 0.1	C > 0.1	Average±SD	$C_{max}$	C < 0.1	C > 0.1	Average±SD	$C_{max}$
Cymoxanil	0	0	-	-	2	0	$0.069 \pm 0.028$	0.088	3	1	0.076±0.022	0.102
Pyrimethanil	4	2	$0.075 \pm 0.069$	0.172	10	2	$0.046 \pm 0.041$	0.113	10	2	$0.059 \pm 0.040$	0.140
Cyprodinil	3	1	$0.262 \pm 0.480$	0.981	11	2	$0.071 \pm 0.037$	0.155	9	1	$0.056 \pm 0.035$	0.121
Iprovalicarb	0	0	-	-	2	0	$0.036 \pm 0.009$	0.042	0	0	-	-
Metalaxyl	5	3	0.421±0.649	1.848	13	2	$0.092 \pm 0.162$	0.666	13	3	0.139±0.333	1.353
CGA 92370	3	4	3.968±7.305	19.97	8	2	$0.106 \pm 0.094$	0.358	6	1	$0.070 \pm 0.036$	0.130
Penconazole	6	3	0.113±0.142	0.476	11	4	$0.066 \pm 0.041$	0.153	18	5	$0.063 \pm 0.069$	0.281
Myclobutanil	2	4	$1.603 \pm 2.244$	5.068	11	0	$0.041 \pm 0.024$	0.082	14	2	$0.065 \pm 0.054$	0.195
Triadimenol	3	4	4.049±9.491	25.52	5	3	$0.076 \pm 0.058$	0.173	9	5	$0.112 \pm 0.171$	0.682
Flutriafol	3	0	$0.016 \pm 0.015$	0.034	4	1	$0.052 \pm 0.064$	0.166	6	1	$0.044 \pm 0.039$	0.110
Tebuconazole	1	3	$1.319 \pm 1.337$	3.023	15	3	$0.061 \pm 0.043$	0.160	14	1	$0.055 \pm 0.035$	0.165
Kresoxim-methyl	2	2	$0.120 \pm 0.085$	0.237	2	9	$0.144 \pm 0.084$	0.362	8	5	$0.084 \pm 0.043$	0.151
Nuarimol	0	0	-	-	0	2	$0.201 \pm 0.016$	0.212	4	2	$0.070 \pm 0.041$	0.120
Benalaxyl	1	1	0.173±0.182	0.302	3	3	$0.091 \pm 0.054$	0.149	6	0	0.036±0.013	0.045
Azoxystrobin	0	2	0.271±0.026	0.289	1	0	0.024	0.024	0	0	-	-
Trifloxystrobin	1	0	0.022	0.022	3	0	$0.037 \pm 0.022$	0.061	9	0	$0.043 \pm 0.018$	0.077
Dimethomorph	1	2	0.196±0.119	0.320	0	0	-	-	0	0	-	-
Cyproconazole	6	1	$0.055 \pm 0.037$	0.107	15	2	$0.053 \pm 0.036$	0.154	14	1	$0.052 \pm 0.028$	0.109

Principal component matrix with the loadings to each of the PCs.

			Ground	l waters			Surface waters					
Variable	PC1	PC2	PC3	PC4	PC5	PC6	PC1	PC2	PC3	PC4	PC5	
Cymoxanil	0.056	0.050	0.075	-0.183	0.480	0.007	0.454	0.148	0.634	-0.241	0.061	
Pyrimethanil	0.506	0.111	0.045	-0.110	0.514	-0.202	0.664	0.010	0.472	-0.371	0.241	
Cyprodinil	0.573	-0.151	-0.319	-0.389	0.244	-0.092	0.701	-0.102	0.536	-0.132	0.052	
Iprovalicarb	0.320	0.540	0.101	0.097	-0.397	-0.046	-0.108	0.363	0.212	0.536	-0.048	
Metalaxyl	0.326	-0.005	-0.372	-0.060	-0.174	0.635	0.374	0.652	-0.461	0.111	-0.097	
CGA 92370	0.542	-0.373	-0.037	0.655	0.108	-0.018	0.303	0.267	-0.380	0.069	0.746	
Penconazole	0.590	0.728	-0.070	0.039	-0.118	-0.038	0.644	0.433	-0.271	-0.146	-0.249	
Myclobutanil	0.669	-0.382	0.212	-0.038	-0.231	-0.211	0.600	0.387	-0.388	-0.252	-0.123	
Triadimenol	0.585	-0.243	-0.558	-0.295	-0.068	0.242	0.345	-0.014	-0.330	0.450	0.665	
Flutriafol	0.415	0.724	-0.129	0.097	-0.089	0.019	0.544	-0.467	-0.028	0.388	-0.149	
Tebuconazole	0.724	-0.463	-0.019	-0.212	-0.160	-0.213	0.660	-0.258	0.115	0.088	0.282	
Kresoxim-methyl	0.575	0.421	0.185	-0.054	0.247	0.075	0.645	-0.347	0.169	0.148	0.111	
Nuarimol	0.083	0.053	0.413	-0.127	0.297	0.581	0.652	-0.170	-0.458	0.274	-0.367	
Benalaxyl	0.489	-0.163	0.536	-0.034	0.193	0.050	0.436	-0.396	0.048	0.085	-0.233	
Azoxyxtrobin	0.302	-0.189	-0.219	0.781	0.256	0.134	0.157	0.534	0.485	0.552	-0.132	
Trifloxystrobin	0.503	0.714	-0.001	0.062	0.029	-0.126	0.648	0.410	-0.050	-0.424	-0.215	
Dimethomorph	0.785	-0.461	0.033	0.007	-0.147	-0.039	0.037	0.450	0.377	0.686	-0.117	
Cyproconazole	0.327	-0.139	0.600	-0.011	-0.374	0.240	0.379	-0.603	-0.252	0.308	-0.230	
% Variance	25.38	16.25	8.55	7.88	7.07	5.80	25.71	14.48	13.03	11.77	8.806	

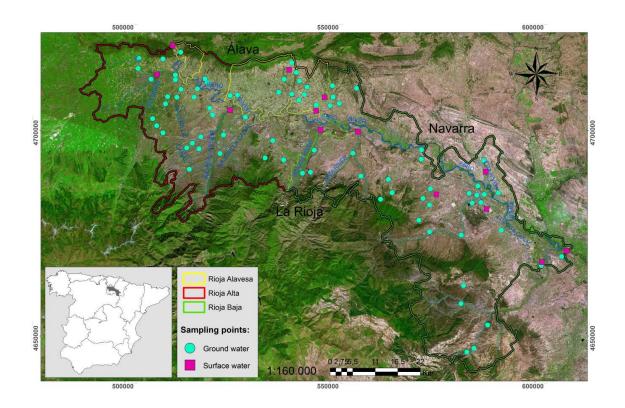


Figure 1

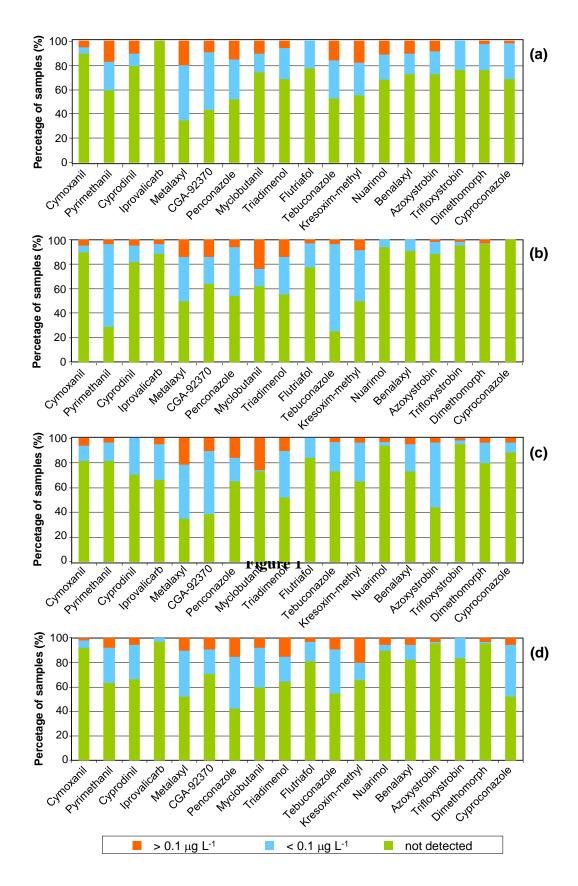


Figure 2

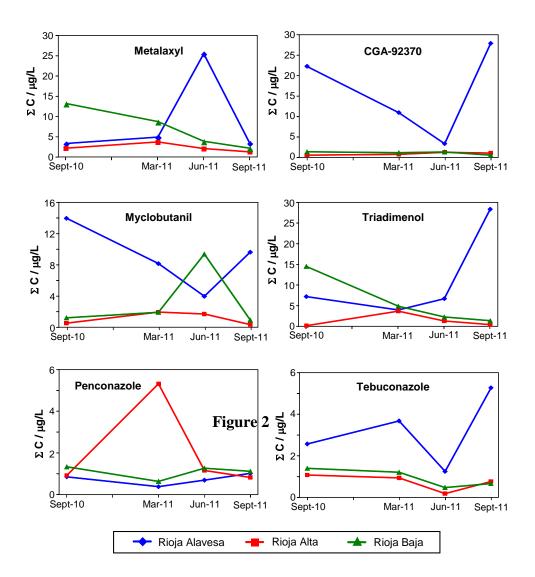


Figure 3

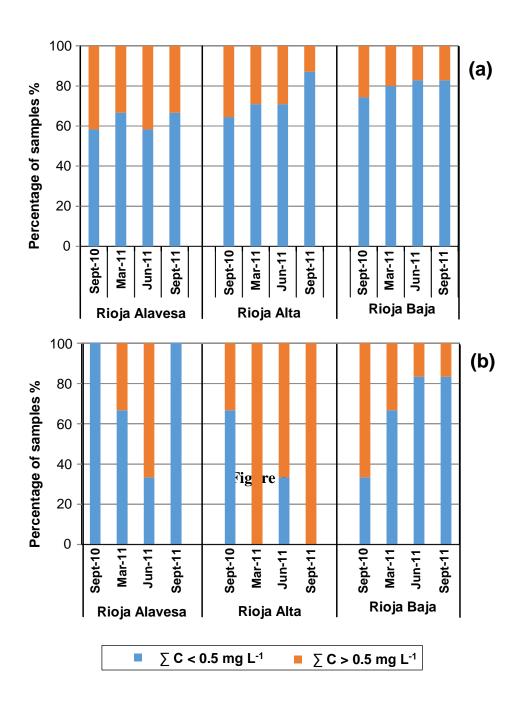


Figure 4.

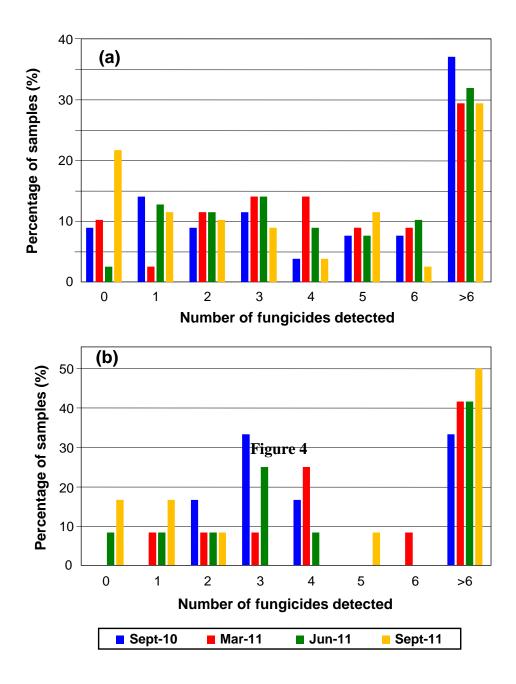


Figure 5.

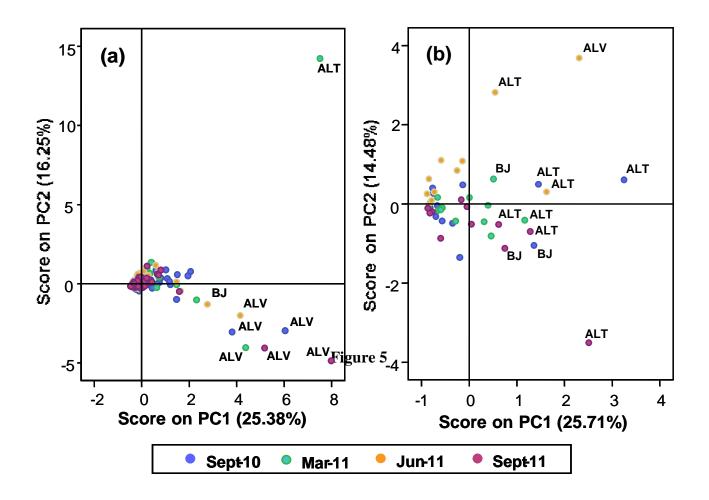


Figure 6

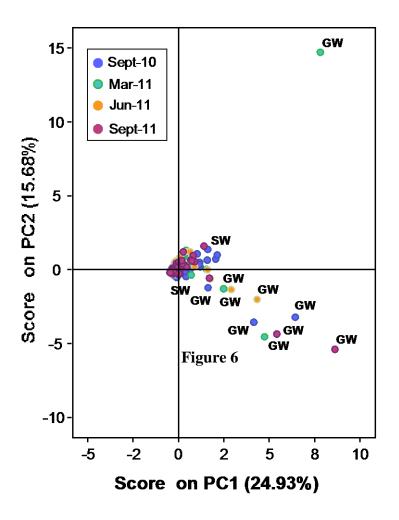


Figure 7

## **Supplementary Material**

# INTRA-ANNUAL TRENDS OF FUNGICIDES RESIDUES IN WATERS FROM VINEYARD AREAS IN LA RIOJA REGION OF NORTHERN SPAIN

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Nº Pages: 7

Nº Tables: 4

Pesticide	Water Solubility $(mg L^{-1})$	GUS index	Log Kow	DT50	Application Dose <sup>a</sup> (g ha <sup>-1</sup> )
Cymoxanil	780	0.34	0.67	1.4	90-120
Pyrimethanil	121	2.65	2.84	55	1750-2000
Cyprodinil	13	1.2	4.0	53	375
Iprovalicarb	17.8	2.35	3.2	10.5	120-150
Metalaxyl	8400	2.11	1.65	42	200-300
CGA 92370	-	-	-	-	-
Penconazole	73.0	1.51	3.72	117	25-75
Myclobutanil	132	3.54	2.89	365	62
Triadimenol	72.0	3.75	3.18	136.7	100-250
Flutriafol	95.0	5.29	2.3	1587	125
Tebuconazole	36.0	2.0	3.7	365	125-250
Kresoxim-methyl	2.0	1.82	3.4	0.87	<300
Nuarimol	26	3.52	3.18	344	150
Benalaxyl	28.6	0.51	3.54	75	150-210
Azoxystrobin	6.0	2.53	2.5	84.5	250
Trifloxystrobin	0.61	0.53	4.5	2	75-180
Dimethomorph	28.95	2.56	2.68	56.7	1500
Cyproconazole	93	3.25	3.09	142	<1

Table S1. Common names, physicochemical properties and application dose of fungicides selected for the study.

<sup>a</sup> Data taken from PPDB (2015) and De Liñán and De Liñán (2008).

Sampling point	Water type	Water depth (m)	Characteristics of the	e area
	water type	water depuir (III)	Crops cultivated	Watering
Rioja Alavesa				
ALV-G1	Groundwater	1-2	Vineyards	Yes
ALV-G2	Groundwater	1-2	Vineyards	Yes
ALV-G3	Groundwater	<5	Cereals and vineyards	No
ALV-G4	Groundwater	Spring	Vineyards	No
ALV-G5	Groundwater	Uptake	Vineyards	No
ALV-G6	Groundwater	Spring	Vineyards	No
ALV-G7	Groundwater	Spring	Vineyards and olives	No
ALV-G8	Groundwater	<5	Vineyards	Yes
ALV-G9	Groundwater	3	Vineyards and cereals	No
ALV-G10	Groundwater	Spring	Vineyards	No
ALV-G11	Groundwater	<5	Vineyards	No
ALV-G12	Groundwater	Spring	Vineyards and cereals	No
ALV-S1	Surface water (Moreda river)	-	Vineyards and olives	No
ALV-S2	Surface water (Oyón river)	-	Vineyards	-
ALV-S3	Surface water (Viñaspre river)	-	Vineyards and orchard	-
Rioja Alta	× ± /		•	
ALT-G1	Groundwater	Spring	Vineyards and cereals	No
ALT-G2	Groundwater	Uptake	Vineyards and cereals	No
ALT-G3	Groundwater	Uptake	Vineyards and cereals	No
ALT-G4	Groundwater	Spring	Vineyards and cereals	No
ALT-G5	Groundwater	Uptake	Cereals, vineyards and beet	No
ALT-G6	Groundwater	8	Cereals, vineyards and beet	No
ALT-G7	Groundwater	<5	Vineyards, cereals and beet	No
ALT-G8	Groundwater	2-3	Vineyards and cereals	No
ALT-G9	Groundwater	Spring	Vineyards and cereals	No
ALT-G10	Groundwater	Uptake	Vineyards	No
ALT-G11	Groundwater	5-10	Vineyards and cereals	No
ALT-G12	Groundwater	<5	Vineyards and fruit trees	No
ALT-G12	Groundwater	Uptake	Vineyards	No
ALT-G13	Groundwater	5-10	Vineyards and cereals	No
	Groundwater		Vineyards, cereals and fruit	NO
ALT-G15	Groundwater	<5	trees	Sometimes
ALT-G16	Groundwater	Uptake	Cereals and vineyards	No
ALT-G17	Groundwater	Uptake	Vineyards	No
ALT-G18	Groundwater	Uptake	Vineyards and orchards	Si
ALT-G19	Groundwater	Uptake	Vineyards and cereals	No
ALT-G20	Groundwater	<5	Vineyards and cereals	Yes, drip
ALT-G21	Groundwater	Uptake	Vineyards and orchards	Yes
ALT-G22	Groundwater	Spring	Vineyards and orchards	Yes
ALT-G22	Groundwater	5-10	Vineyards	No
ALT-G24	Groundwater	Uptake	Vineyards	No
ALT-G25	Groundwater	3	Vineyards	No
ALT-025 ALT-G26	Groundwater	45	Vineyards and orchards	No
ALT-G20 ALT-G27	Groundwater	Spring	Vineyards and cereals	No
ALT-G27 ALT-G28	Groundwater		-	No
		Spring	Vineyards and orchards	
ALT-G29	Groundwater	5-6	Vineyards	No
ALT-G30	Groundwater	Uptake	Vineyards and cereals	No

**Table S2.** Characteristics of the sampling points monitored in the three subareas studied in La Rioja region.

Sampling point	Water type	Water depth (m)	Characteristics of the area				
Sampling point	Water type	water depth (III)	Crops cultivated	Watering			
ALT-G31	Groundwater	2-3	Vineyards, cereals and potatoes	Yes			
ALT-S1	Surface water (Ebro river)	-	Vineyards, olive and fruits				
ALT-S2	Surface water (Najerilla river)	-	Vineyards				
ALT-S3	Surface water (Oja river)	-	Vineyards				
Rioja Baja							
BJ-G1	Groundwater	Spring	Vineyards	No			
BJ-G2	Groundwater	60	Vineyards	Yes, drip			
BJ-G3	Groundwater	Spring	Fruit trees and orchards	Yes			
BJ-G4	Groundwater	Uptake	Fruit trees	Yes			
BJ-G5	Groundwater	<3	Olive trees	Yes, drip			
BJ-G6	Groundwater	3-4	Vineyards	No			
BJ-G7	Groundwater	Spring	Fruit trees	No			
BJ-G8	Groundwater	2-3	Vineyards	Yes, drip			
BJ-G9	Groundwater	3	Vineyards	Yes, drip			
BJ-G10	Groundwater	4-5	Vineyards	Yes, drip			
BJ-G10 BJ-G11	Groundwater	5	Vineyards and fruit trees	Yes			
BJ-G12	Groundwater	3-4	Vineyards	No			
BJ-G13	Groundwater	<5	Orchards	Yes			
BJ-G14	Groundwater	5-6	Vineyards and olives	No			
BJ-G15	Groundwater	Spring	Vineyards	Yes, drip			
BJ-G16	Groundwater	3-4	Vineyards and cereals	Sometimes			
BJ-G17	Groundwater	17	Fruit trees and orchards	Yes			
BJ-G18	Groundwater	7-9	Vineyards and olives	Yes, drip			
BJ-G19	Groundwater	8-10	Vineyards and olives	Yes, drip			
BJ-G20	Groundwater	> 10	Vineyards	No			
BJ-G21	Groundwater	> 10	Olive trees	No			
BJ-G22	Groundwater	> 10	Orchards	Yes			
BJ-G23	Groundwater	Spring	Vineyards, cereals and olives	No			
BJ-G24	Groundwater	5-10	Vineyards and cereals	No			
BJ-G25	Groundwater	5-6	Orchards	Yes			
BJ-G26	Groundwater	Spring	Vineyards and cereals	No			
BJ-G27	Groundwater	Spring	Vineyards, cereals and olives	No			
BJ-G28	Groundwater	<5	Vineyards	No			
BJ-G29	Groundwater	3-4	Vineyards	Yes			
BJ-G30	Groundwater	Spring	Vineyards	No			
BJ-G31	Groundwater	3-4	Vineyards, cereals, fruit trees	No			
BJ-G32	Groundwater	6-8	Vineyards	No			
BJ-G33	Groundwater	Spring	Vineyards and olives	No			
BJ-G34	Groundwater	6-8	Vineyards	No			
BJ-G35	Groundwater	5-6	Vineyards	Yes, drip			
BJ-S2	Surface water (Lodosa canal)	-	Vineyards and fruit trees	s, unp			
BJ-S3	Surface water (Ebro river)	-	Vineyards				
BJ-S5 BJ-S4	Surface water (Ega river)	-	Vineyards				
BJ-S4 BJ-S5	Surface water (Iregua river)	-	Vineyards				
BJ-55 BJ-S6	Surface water (Leza river)	-	Vineyards				
	Surface water (Villar de		-				
BJ-S7	Arnedo river)	-	Vineyards				

Fungicide	SIM ion m/z	V cone (V)	SIM window	RT (min)	Recovery <sup>a</sup> (%)	RSD (%)	$r^{2}$ (0.1-2.0 $\mu g L^{-1})^{b}$	$LOD^{c}$ (µg L <sup>-1</sup> )	$LOQ^d$ (µg L <sup>-1</sup> )
Cymoxanil	199.2	35	1	8.3	76	13	0.991	0.022	0.069
Cyproconazole	292.2	20	4	18.3	76	13	0.992	0.021	0.059
Azoxystrobin	404.2	25	4	18.3	85	11	0.995	0.017	0.052
Iprovalicarb	321.3	20	5	19.9	86	7	0.995	0.016	0.037
Dimethomorph	388.2	25	5	20.6	82	12	0.995	0.017	0.062
Benalaxyl	326.2	25	6	25.2	89	15	0.992	0.015	0.048
Cyprodinil	226.2	40	6	25.5	73	9	0.994	0.018	0.043
Trifloxystrobin	409.1	20	6	26.9	64	7	0.991	0.016	0.037

**Table S3.** Conditions for analytical determination and quality control parameters of the SPE-LC-MS method applied to the analysis of fungicides in surface and ground waters.

<sup>a</sup> Calculated from the replicated analysis (n = 5) of spiked (0.1  $\mu$ g L<sup>-1</sup>) groundwater samples; <sup>b</sup> Linear calibration range; <sup>c</sup> LOD Detection limit for a signal-to-noise ratio of 3. <sup>d</sup> LOQ Quantification limit for a signal-to-noise ratio of 10

		•	6	e				
Compound	SIM window	RT (min)	Monitored ions (Abundance)	Recovery <sup>a</sup> (%)	RSD <sup>a</sup> (%)	r <sup>2</sup> (0.1-1.5 <sup>b</sup> μg L <sup>-1</sup> )	$LOD^{c}$ (µg L <sup>-1</sup> )	$LOQ^d$ (µg L <sup>-1</sup> )
	willdow	(11111)	Target (m/z) Qualifier Ions (m/z)					
CGA 92370	1	7.3	148 120 (572)/91 (322)	78	17			
Pyrimethanil	3	10.0	198 199	65	14	0.992	0.031	0.078
Metalaxyl	4	12.9	45 206 (326)/132 (299)	108	12	0.997	0.001	0.006
Penconazole	5	16.3	248 159 (851)/161 (577)	83	14	0.996	0.002	0.005
Triadimenol	5	16.4	112 128 (906)/168 (745)	77	15	0.993	0.005	0.021
Flutriafol	5	20.6	123 164 (552)/219 (255)	80	17	0.997	0.005	0.013
Myclobutanil	5	21.4	179 82 (451)/150 (449)	83	15	0.999	0.003	0.013
Kresoxim- methyl	5	22.2	116 131 (533)/206 (516)	91	14	0.989	0.016	0.041
Tebuconazole	6	24.6	125 250 (847)/70 (492)	84	17	0.997	0.006	0.022
Nuarimol	6	25.3	107 139 (765)/235 (744)	83	14	0.994	0.124	0.284

**Table S4.** Conditions for analytical determination and quality control parameters of the SPE-GC-MS method applied to the analysis of fungicides in surface and ground waters.

<sup>a</sup> Calculated from the replicated analysis (n = 5) of spiked (0.1  $\mu$ g L<sup>-1</sup>) groundwater samples; <sup>b</sup> Linear calibration range. <sup>b</sup> Lineal range. <sup>c</sup> LOD Detection limit for a signal-to-noise ratio of 3. <sup>d</sup> LOQ Quantification limit for a signal-to-noise ratio of 10

#### References

- De Liñán Vicente C, De Liñán Carral C (2008) Vademecum de productos fitosanitarios y nutricionales. Agrotécnicas S.L., Spain.
- PPDB, Pesticide Properties DataBase (2015) University of Hertfordshire. Publishing in http://sitem.herts.ac.uk/aeru/ppdb/en/index.htm. Accessed April 2015.